CHEMICAL CONSTITUENTS AND EVALUATION OF ANTIPROLIFERATIVE AND ANTI-INFLAMMATORY ACTIVITIES FROM Psychotria schlechtendaliana (Rubiaceae)

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Phytochemical study of Psychotria schlechtendaliana (Rubiaceae) aerial parts resulted in the isolation of alkaloids 4-N-oxide harmaine (1) and strictosidine acid (2), and the terpenoids sitosterol, estigmastanol, α-amyrin, β-amyrin and betulinic acid. The crude extract (CE), its fractions (hexane FH, chloroform FC, ethyl acetate FEA, hydromethanolic FHM, alkaloidal chloroform FCOH, alkaloidal aqueous FAq) and majority alkaloid (I) were investigated for their antiproliferative potential against nine human tumor cell lines and one non-tumoral human cell line (HaCat). CE, FH and FEA fractions exhibited strong growth inhibition for ovary cells (OVCA-3, GI50 = 5.89; 1.36 and 6.59 μg mL−1, respectively) and FC, FH and FAm fractions showed potent activity on the growth of leukemia cell lines (K562, GI50 = 1.92; 7.23 and 8.81 μg mL−1, respectively). Compound I exhibited selective antiproliferative activity to breast cancer (MCF-7, GI50 = 32.7 μg mL−1) and was non-toxic to HaCat cells. To evaluate the anti-inflammatory effect, models of ear edema induced by croton oil and the enzyme myeloperoxidase (MPO) were used. FH and FEA factions exhibited anti-inflammatory effect and reduce ear edema compared to the control group. These fractions showed results superior to those exhibited by indomethacin (75.1%), FH (97.5%) and FC (95.5%) fractions.

Keywords: alkaloids; terpenoids; Psychotria; antiproliferative activity; anti-inflammatory activity.

INTRODUCTION

Psychotria is the largest genus within Rubiaceae, comprising approximately 1700 species distributed in the tropical and subtropical regions of the world.1,2 Several South American Psychotria species are used as medicinal plants to treat different diseases, including gastrointestinal disorders, and has antibacterial, antiviral, antifungal, anti-inflammatory, and anticancer action. These different biological activities are associated with substances of the classes of alkaloids, iridoids, terpenoids, and flavonoids recurrently described in the genus.3,4 Psychotria schlechtendaliana (Müll. Arg.) Müll. Arg., synonym of Palicourea divaricata Schltdl and Uragoga schlechtendaliana (Müll. Arg.) Kuntze, is an endemic species from Brazil Northeast in the caatinga (semi-arid) and Atlantic Forest area.5 As part of our continuing work on the Rubiaceae, in this study we reported the isolation of β-carbolines alkaloids and triterpenes and, in addition, antiproliferative, and anti-inflammatory activities of P. schlechtendaliana. This is the first chemical and pharmacological study of this species.

EXPERIMENTAL

General experimental procedures

The NMR analysis was performed out on a Bruker Avance III HD (500 MHz or 300 MHz for 1H and 125 MHz or 75 MHz for 13C), in CDCl3 and/or CD3OD (Merck, Barueri-SP, Brasil), using TMS as an internal standard. The chemical shifts (δ) were expressed in ppm and coupling constants (J) in Hz. Column chromatography (CC) was performed on silica gel 60 (0.063-0.200 mm, Merck) and Sephadex LH-20 (Sigma-Aldrich, Barueri-SP, Brasil). Thin layer chromatography (TLC) was performed on silica gel 60 (5.0 × 20 cm × 0.20 mm). The organic solvents used in the methodologies were n-hexane (95.8%), chloroform (99.5%), ethyl acetate (99.5%), n-butanol (99.4%), and methanol (99.8%) from Synth (São Paulo, Brazil).

Plant material

Aerial parts of de P. schlechtendaliana were collected in Private Reserve of the National Patrimony of Serra Bonita (PRNP-Serra Bonita), on Camacan, South of Bahia, Brazil (Bapeba Trail: 15º23’30” S, 39º33’55” W), in December of 2013, identified by Prof. Dr. André Márcio Amorim and a voucher specimen (CEPEC 141.213) was deposited in the herbarium of the State University of Santa Cruz. P. schlechtendaliana was registered in the Sistema Nacional de Gestão do Patrimônio Genético e Conhecimento Tradicional Associado (SisGen) at the number A980835.

Extraction and isolation

The dried and ground aerial parts of P. schlechtendaliana (273 g, 16 mesh) were degreased with n-hexane and extracted with methanol (18 × 500 mL, 72 h), obtaining the crude methanolic extract (CE, 44.52 g) after concentration on a rotary evaporator (40 °C). Part of the CE (19.60 g) was suspended in methanol:water (1:1 v/v) and subjected to acid-base extraction with hydrochloric acid solution (10% v/v) and CHCl3, obtaining the chloroform-acid fraction (FCH, 6.02 g). Acid solution residue was basified with NH4OH to pH 10 and extracted again with CHCl3, providing the alkaloidal chloroform (FCOH, 90.0 mg) and the aqueous fractions (FAq, 10.0 g). Fraction FAq (9.0 g) was suspended in water (100 mL) and partitioned with ethyl acetate.
and n-butanol, providing the ethyl acetate (FEA1, 95.0 mg), butanol (FB1, 275.7 mg) and aqueous fractions (FAq1, 8.0 g), respectively. Compound 1 (8.0 mg) was obtained from fraction FCOH (80.0 mg) by a preparative TLC, eluting with chloroform:methanol (8:2) and NH₄OH drops, followed an extraction of the material with CHCl₃:MeOH (1:1). Compound 2 (5.0 mg) was isolated from FB1 fraction (275.7 mg) by successive fractionation by CC on silica gel 60 (14.56 g, Θ = 1.5 cm, h = 21 cm), with CHCl₃:MeOH in the ratio of 10:0 to 10:0 (with NH₄OH drops). Another part of CE (9.62 g) was suspended in MeOH:H₂O (1:1) and partitioned with hexane, chloroform and ethyl acetate. This procedure resulted in hexane (952.9 mg), chloroform (FC, 615.0 mg), ethyl acetate (FEA, 134.0 mg), and hydromethanol fractions (FHM, 6.39 mg). The FH fraction (400 mg) was subjected to chromatographic column silica gel 60 (18.00 g, Θ = 1.6 cm, h = 30 cm), eluted in Hex:EtOAc in an increasing polarity gradient, 10 subfractions (FH1-FH10) were obtained and in FH3 (3.0 mg) a white solid was provided which was named by substances 3 and 4. FC fraction (600.0 mg) was treated on silica gel CC (21.00 g, Θ = 2.0 cm, h = 25 cm), eluted in hexane, ethyl acetate and methanol in different proportions, in increasing order of polarity. From this, 27 fractions were obtained (FC1-FC27), and fraction FC15 (141.0 mg) was purified on silica gel CC (9.62 g, Θ = 1.45 cm, h = 24 cm) using the CHCl₃:MeOH solvent system in a gradient of 10:0 at 0:10, giving a mixture of substances 5, 6 and 7 as white powder.

Strictosidinic acid (2) 1H NMR (500 MHz, CD₃OD) δ 4.47 (d, 1H, J = 11.9 Hz, H-3), 3.72 (m, 2H, H-5), 7.45 (d, 1H, J = 7.6 and 7.9 Hz, H-10), 7.12 (dd, 1H, J = 7.6 and 8.1 Hz, H-11), 7.31 (d, 1H, J = 8.1 Hz, H-12), 2.12 (dd, 1H, J = 13.8, 11.4 and 4.2 Hz, H-14a), 2.27 (dd, 1H, J = 13.8, 13.0 and 3.2 Hz, H-14b), 2.98 (m, 1H, H-15), 7.62 (brs, 1H, H17), 5.21 (d, 1H, J = 9.6 Hz, H-21), 4.82 (d, 1H, J = 7.9 Hz, H-1’), 3.24 (dd, 1H, J = 10.4, 2.4 Hz, H-2’), 3.45 (dd, 1H, J = 7.2, 2.4 Hz, H-3’), 3.25 (dd, 1H, J = 10.4, 7.2 Hz, H-4’), 3.41 (t, 1H, J = 9.0 Hz, H-5’), 3.68 (dd, 1H, J = 11.9, 7.2 Hz, H-6’a), 4.01 (dd, 1H, J = 11.9, 1.9 Hz, H-6’b); 13C NMR (125 MHz, CD₃OD) δ 130.6 (C-2), 52.6 (C-3), 43.1 (C-5), 19.7 (C-6), 107.4 (C-7), 127.6 (C-8), 119.2 (C-9), 129.0 (C-10), 123.5 (C-11), 112.4 (C-12), 138.3 (C-13), 35.2 (C-14), 33.8 (C-15), 112.8 (C16), 154.2 (C-17), 119.2 (C-18), 136.2 (C-19), 45.8 (C-20), 96.9 (C-21), 175.5 (C-22), 100.5 (C-1’), 74.9 (C-2’), 78.1 (C3’), 72.0 (C-4’), 78.9 (C-5’), 63.2 (C-6’).

In vitro assays

Cell lines and culture conditions

For in vitro antiproliferative screening, the nine human tumor cell lines were tested, being glioma (U-251), breast (MCF-7), ovary, with multiple drug resistance phenotype (NCI-ADR/RES), kidney (786-O), lung, non-small cell type (NCI-H40), prostate (PC-3), ovary (OVCA-03), colon (HT-29), leukemia (K562). The Frederick Cancer Research and Development Center (National Cancer Institute, Frederick, MA, USA) provided these cell lines. The immortalized keratinocyte non-tumor cell line (HaCat) was donated by Prof. Dr. Ricardo Della Coletta, FOP/UNICAMP. All human cells were cultivated in a 75 cm² flask (Corning, USA) in complete medium (Roswell Park Memorial Institute (RPMI) medium 1640 (GIBCO, NY, USA) supplemented with 5% fetal bovine serum (FBS) (Vitrocell, SP, BR) and penicillin: streptomycin (1000 U mL⁻¹:1000 μg mL⁻¹, 10 mL L⁻¹ in RPMI 1640, Vitrocell, SP, BR).]

Antiproliferative assay

The samples (10 mg; CE, FH, FC, FEA, FHM, FCOH, FAq and compound 1) were diluted in dimethyl sulfoxide (DMSO) followed by successive dilutions in a complete medium to give final concentrations of 0.25; 2.5; 25 and 250 μg mL⁻¹ and incubated at 37 °C in a 5% CO₂ atmosphere in a humid environment for 48 h. Doxorubicin hydrochloride 0.4% m/v (Europharma Green Bay, WI) was used as a positive control. Another microplate containing all cell lines was also prepared and use to establish the cell quantities at sample addition (time zero, T₀). Subsequently, cells were fixed with 50 μL of 50% trichloroacetic acid (TCA) and cell proliferation was determined by spectrophotometry (540 nm) of the protein content using the sulfurhodamine B assay. Results were plotted by correlating drug concentration and proliferation for each cell type. The values of GI₀ (sample concentration required to 50% inhibition of cell proliferation) by nonlinear regression sigmoid type were established, using Origin 7.5 software.

In vivo evaluations

Animals

The anti-inflammatory activity assay was performed using male Swiss mice, weighing 30–40 g kept in collective cages (five animals) in an environment-controlled humidity (40-60%) and temperature (22 ± 2 °C), under a 12/12 h light/dark cycle, with free access to food and water. The protocols for these experiments were approved by the Committee of Ethics in Animal Experimentation of the State University of Maringa (CEUA/UEM 9804/2016).

Croton oil-induced ear edema

The edema was induced by the application of 20 μL of CO (200 μg) diluted in 70% acetone (acetone/water 3:7 v/v) at the inner surface of the left ear (LE) of mice. Crodust extract (20 μL, 2.5 mg ear⁻¹) and fractions (FH, FC, FEA and FHM; 20 μL, 2.5 mg ear⁻¹) were applied to the left ear, and the same volume of solvent was applied to the right ear as control. Indomethacin was used as a positive control (1.0 mg ear⁻¹). After 6 h, the animals were anesthetized, euthanized, and each ear was perforated with a metal punch to provide a 6 mm diameter disc. Edema was assessed by subtracting the weight of the disc from the right control ear from the weight of the disc from the left treated ear, thus determining the percentage of edema inhibition. The percentage of edema inhibition was determined according to Bracht et al.

Myeloperoxidase activity (MPO)

The myeloperoxidase activity (MPO) was assessed according to Boller et al. Ear tissue was placed in 50 mmol L⁻¹ potassium phosphate buffer, pH 6.0, containing 0.5% hexadecyltrimethylammonium bromide (1 mL 50 mg⁻¹ tissue, Sigma-Aldrich) in Potter homogenizer. The homogenate was centrifuged for 5.0 min at 2500 rpm and 25 °C and supernatant was added to a 96-well microplate, followed by the addition of 200 mL of the buffer solution containing o-dianisidine dihydrochloride (16.7 mg, Sigma-Aldrich), distilled water (90 mL), potassium phosphate buffer (10 mL), and H₂O₂ 1% (50 mL). The reaction was stopped by the addition of 30 μL of 1.46 mol L⁻¹ sodium acetate, pH 3.0 and MPO activity was determined colorimetrically using a microplate reader to measure absorbance at 450 nm and expressed as OD per tissue.

Statistical analysis

For the results of anti-inflammatory activities (n = 6) the data were expressed as mean values ± SEM (standard error of the mean) and tested with analysis of variance (ANOVA) by Tukey test, being considered significant p < 0.05.
RESULTS AND DISCUSSION

Chemical characterization

The phytochemical investigation of the aerial parts of Psychotria schlechtendaliana afforded seven compounds (Figure 1): the alkaloids 4-N-oxide-harmine (1),
stri恭喜dicin acid (2),
steroids sitosterol (3) and stigmasteryl (4), and the triterpenes α-amininy (5),
β-amininy (6) and betulinic acid (7). The chemical structures were characterized by 1H, 13C, and 2D NMR analysis, followed by comparison with the literature.

![Figure 1. Molecular structure of compounds isolated from aerial parts of P. schlechtendaliana](image)

The 1H, 13C and 2D NMR spectra of compound 1 showed six aromatic hydrogens at δH 8.45 (d, J 6.2 Hz, H-6), 8.29 (dt, J 8.1, 1.0 Hz, H-9), 8.30 (d, J 6.2 Hz, H-5), 7.76 (m, H-11), 7.75 (m, H-12) and 7.45 (ddd, J 8.2, 6.2, 1.6 Hz, H-10). The singlet at δH 3.05 (s, CH3) was characterized as a methyl group in the position H-14. By the heteronuclear single quantum correlation (HSQC) experiment, the correlations of all hydrogens with their respective carbons were established (Table 1). The HMBC spectrum showed the correlations between the H-5/C-2/C-6, H-6/C-7, and H-10/C-12 signals belonging to the indolic nucleus. The spectral data were compared with the literature for the harmine alkaloid.

![Table 1. Spectroscopic data of 1H/HSQC (300 MHz) and 13C (75.45 MHz) NMR of alkaloid 4-N-oxide-harmine (1) in CDCl3](image)

<table>
<thead>
<tr>
<th>Position</th>
<th>δC</th>
<th>δH/HSQC</th>
</tr>
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<tbody>
<tr>
<td>2</td>
<td>133.9</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>145.1</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>130.8</td>
<td>8.30 (d, J 6.2)</td>
</tr>
<tr>
<td>6</td>
<td>116.3</td>
<td>8.45 (d, J 6.2)</td>
</tr>
<tr>
<td>7</td>
<td>135.9</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>121.9</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>124.1</td>
<td>8.29 (d, J 8.2, 1.0)</td>
</tr>
<tr>
<td>10</td>
<td>122.9</td>
<td>7.45 (ddd, J 8.2, 6.2, 1.6)</td>
</tr>
<tr>
<td>11</td>
<td>132.7</td>
<td>7.76 (m)</td>
</tr>
<tr>
<td>12</td>
<td>113.8</td>
<td>7.75 (m)</td>
</tr>
<tr>
<td>13</td>
<td>140.1</td>
<td></td>
</tr>
<tr>
<td>14</td>
<td>16.8</td>
<td>3.05 (s)</td>
</tr>
</tbody>
</table>

δ in ppm; J in Hz.

other monoterpenoid indol alkaloids and pyrrolidine indol alkaloids.21-23 Psychotria is a genus taxonomically complex and due to morphological characters and geographical distribution. It is recognized three subgenera: Psychotria (pantropical), Tetrameraceae (includes some species from Africa and Madagascar) and Heteropsychotria (includes the remainder of the species of Psychotria in the neotropics).24

Several alkaloid types are described on the Psychotria genus and the presence of different profiles of these was observed in the different subgenera. Alkaloids of polypeptide type are frequent in Psychotria subgenus, while indole monoterpenoid alkaloids are related to the Heteropsychotria subgenus, especially the species collected in Brazil.25 The presence of compounds 1 and 2 in P. schlechtendaliana corroborates this observation.

In a recent study, we relate the isolation of strictosidinic acid (2) of Palicourea minitiflora and, in the evaluation of anti-inflammatory activity was observed that this substance presented a low effect of inhibiting the formation of edema in anti-inflammatory activity, however promoted a reduction of 81.9% (1.25 mg ear-1) in the MPO activity. In the assay of antiproliferative activity this alkaloid did not present any effect on the antitumor cell lines at the highest tested concentration.

Antiproliferative assay

The extract and their fractions exhibited activity for different tumor cells (Table 2), according to the parameters established by N'Da and Smith26 for growth inhibition (GI50). Crude extract (CE) exhibited potent activity cytostatic non-selective for ovarian line (OVCAR-3, GI50 = 5.89 µg mL-1) and moderate activity for leukemia (K562, GI50 = 22.22 µg mL-1). Fraction FH showed results against ovary (OVCAR-03, GI50 = 1.36 µg mL-1), leukemia (K562, GI50 = 7.23 µg mL-1), glioma (U251, GI50 = 25.84 µg mL-1), breast (MCF-7, GI50 = 25.72 µg mL-1), ovarian (NICI-ADR/RES, GI50 = 29.37 µg mL-1), kidney (786-O, GI50 = 26.01 µg mL-1), lung (NICI-H40, GI50 = 30.12 µg mL-1) and prostate (PC-3, GI50 = 24.35 µg mL-1) cell lines. In addition, for the non-tumor cell line (HaCat, GI50 = 28.64 µg mL-1), Fraction FC indicated potent activity for leukemia (K562, GI50 = 7.23 µg mL-1), and moderate activity for ovary (OVCAR-03, GI50 = 25.00 µg mL-1). FEA fraction was active for ovary (OVCAR-03, GI50 = 6.59 µg mL-1) and leukemia (K562, GI50 = 24.50 µg mL-1) cell lines. The alkaloid fraction (FCOH)
Table 2. Cytotoxicity effects of the crude extract, fractions, and compound 1 in the different cells

<table>
<thead>
<tr>
<th>Cell lines</th>
<th>Dox</th>
<th>CE</th>
<th>FH</th>
<th>FC</th>
<th>FEA</th>
<th>FHM</th>
<th>FCOH</th>
<th>FAq</th>
<th>I</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glioma (U-251)</td>
<td>&lt;0.025</td>
<td>72.20</td>
<td>25.84</td>
<td>55.00</td>
<td>30.33</td>
<td>&gt; 250</td>
<td>25.92</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Breast (MCF-7)</td>
<td>0.071</td>
<td>65.08</td>
<td>25.72</td>
<td>78.53</td>
<td>62.87</td>
<td>&gt; 250</td>
<td>241.5</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Ovarian (NCI-ADR/RES)</td>
<td>0.15</td>
<td>113.1</td>
<td>29.37</td>
<td>103.4</td>
<td>208.9</td>
<td>&gt; 250</td>
<td>244.7</td>
<td>250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Kidney (786-O)</td>
<td>0.11</td>
<td>73.57</td>
<td>26.01</td>
<td>49.87</td>
<td>52.51</td>
<td>&gt; 250</td>
<td>44.92</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Lung (NCI-H40)</td>
<td>&lt;0.025</td>
<td>246.3</td>
<td>30.12</td>
<td>55.50</td>
<td>75.41</td>
<td>&gt; 250</td>
<td>11.61</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Prostate (PC-3)</td>
<td>0.040</td>
<td>85.49</td>
<td>24.35</td>
<td>55.24</td>
<td>55.44</td>
<td>&gt; 250</td>
<td>158.0</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Ovary (OVCAR-03)</td>
<td>&lt;0.025</td>
<td>5.89</td>
<td>1.36</td>
<td>25.00</td>
<td>6.59</td>
<td>&gt; 250</td>
<td>35.29</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Colon (HT-29)</td>
<td>0.045</td>
<td>180.0</td>
<td>33.39</td>
<td>218.1</td>
<td>31.09</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>Leukemia (K562)</td>
<td>&lt;0.025</td>
<td>22.22</td>
<td>7.23</td>
<td>1.92</td>
<td>24.50</td>
<td>&gt; 250</td>
<td>12.66</td>
<td>8.81</td>
<td>&gt; 250</td>
</tr>
<tr>
<td>HaCat</td>
<td>0.026</td>
<td>65.79</td>
<td>28.64</td>
<td>38.99</td>
<td>38.99</td>
<td>&gt; 250</td>
<td>157.8</td>
<td>&gt; 250</td>
<td>&gt; 250</td>
</tr>
</tbody>
</table>

Values were determined through nonlinear regression analysis using Origin 7.5. Dose range evaluated 0.25-250 µg mL⁻¹. Assessed by the SRB assay. GI₅₀: concentration required inhibiting 50% of cell proliferation. Dox: doxorubicin (positive control); CE: crude extract; FH: hexane; FC: chloroform; FEA: ethyl acetate; FHM: hydromethanol; FCOH: alkaloidal chloroform; FAq: aqueous fraction. Inactive: GI₅₀ > 100 µg mL⁻¹; weak activity: 30 < GI₅₀ < 100 µg mL⁻¹; moderate activity: 10 < GI₅₀ < 30 µg mL⁻¹; potent activity: GI₅₀ < 10 µg mL⁻¹.²⁹

![Figure 2. Effect of topical treatment with P. schlechtendaliana extract and fractions. (2.5 mg ear⁻¹) on-ear edema, 6 h after croton oil (CO) application in the inner surface of the ear in mice (n = 6 animals/group). Indo (1 mg ear⁻¹) were use as anti-inflammatory drugs (positive control). V: vehicle (20 µL acetone 70% v:v); CO: croton oil (inflamed control group); Indo: indomethacin; CE: crude extract; FH: hexane; FC: chloroform; FEA: ethyl acetate; FHM: hydromethanol. The data was express as the mean ± SEM for each group. Different letters indicate the statistical difference between groups (p < 0.05, ANOVA followed by Tukey’s test).

Psychotria species have been use in folk medicine as anti-inflammatory agents, alkaloids and terpenoids are important compounds that lead to these results. Some studies were developed looking for applications such as anti-inflammatory drugs. The inhibition of nitric oxide (NO) caused by compounds isolated from P. prainii,³¹ P. nuda,³² and P. capensis³³ reinforces the responses of the extracts, in addition to indicating the need for further studies with the genus. The evaluation of the in vivo anti-inflammatory capacity carrageenan-induced paw edema in rats indicated that the fruits and stems of P. nilgiriensis showed significant inhibition.³⁴

Triterpenoids, in general, have several pharmacological applications and respond efficiently to various biological assays. α-Amyrin (5), β-amyrin (6), betulenic acid (7) and β-sitosterol (4) are compounds that have numerous descriptions in natural products, in addition to their pharmacological potential been consolidated in the literature. Research reports the anti-inflammatory response of these substances by different models in vitro and in vivo. The significant paw edema inhibitory activity was observed by β-amyrin (6),³³,³⁶ β-Sitosterol (4), α-amyrin (5) showed significant
anti-edematogenic activity in comparison with that of indomethacin in mouse ear. The inhibitory effects against TPA-induced edema as well as against carrageenan-induced edema for 5 and 6 have also been reported. These triterpenoids are also antitumor for different cells. The compounds 5 and 6 exerts anticancer efficacy by several mechanisms such as proliferation inhibition, enhancing apoptosis and growth receptor modulation, betulinic acid (7) is described as a potent selective inhibitor of human melanoma in vitro and in vitro and its anti-inflammatory activity was shown in different models of inflammation. Antitumor activities also have been observed for sitosterol and stigmasterol steroids. Therefore, the presence of these metabolites could explain the positive results presented by apolar fractions in anti-inflammatory and antiproliferative assays.

CONCLUSION

Our findings reveal that Psychotria schlechtendaliana has compounds with antiproliferative and anti-inflammatory activity. Compound 1, the alkaloid 4-N-oxide harmaine exhibited selective antiproliferative activity to human breast cancer and was non-toxic to HaCaT cells, showing a promising source in the search for new drugs. The harmane-f-carboline alkaloid and its derivatives have shown several pharmacological activities such as antiangiogenesis, antidepressant, antiplatelet, spasmylocytic, antiinflammatory, acetylcholinesterase, and antiplatelet, spasmolytic, antidiabetic, acetylcholinesterase, and antidiabetic activities as well as against carrageenan-induced edema for 5 and 6. The inhibitory effects against TPA-induced edema as well as against carrageenan-induced edema for 5 and 6 have also been reported. These triterpenoids are also antitumor for different cells. Therefore, the presence of these metabolites could explain the positive results presented by apolar fractions in anti-inflammatory and antiproliferative assays.

SUPPLEMENTARY MATERIAL

Supplementary data (1H and 13C NMR spectra of compounds 1 and 2) are available free of charge at http://quimicanova.sbq.org.br as PDF file.

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